



Westfälische  
Wilhelms-Universität  
Münster

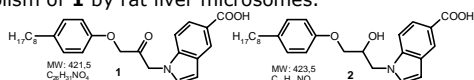
# METABOLISM OF A CYTOSOLIC PHOSPHOLIPASE $A_2\alpha$ INHIBITOR WITH AN ACTIVATED KETO GROUP BY RAT LIVER MICROSOMES

Jörg Fabian, Matthias Lehr

Institute of Pharmaceutical and Medicinal Chemistry, University of Münster  
Hittorfstr. 58-62, 48149 Münster, Germany

## Introduction:

Inhibitors of cytosolic phospholipase  $A_2\alpha$  (cPLA $_{2\alpha}$ ) are assumed to provide a novel therapeutic with application in many inflammatory diseases. Recently we have found that 1-[3-(4-octylphenoxy)-2-oxypropyl]indole-5-carboxylic acid (**1**) is a potent inhibitor of cPLA $_{2\alpha}$  with an activated ketone moiety as pharmacophore (serine trap) [1]. Since it is known that this functional group shows metabolic instability we investigated the metabolism of **1** by rat liver microsomes.



It is important to evaluate the metabolic stability of drug candidates toward CYP-mediated metabolism, and to use it as parameter in identifying and optimizing drug candidates for further development [2].

Though inter-species differences and variations of CYP isoforms are known [3] metabolism by rat liver microsomes give an idea about stability and potential bioavailability of substances.

## Microsomal Incubation:

The liver microsomes of male Sprague-Dawley rats were prepared adapted to published procedures [4,5].

Microsomal incubations were carried out in 0.1 M potassium phosphate buffer pH 7.4 at 37 °C in a shaking water bath and contained rat microsomes (2.25 [mg/ml] protein / Bradford / BSA standards), 3 mM MgCl $_2$  and 1.9 mM NADPH. The final addition of the analyt (20  $\mu$ M) initiated the metabolism which was carried out for 30 min (2h for LC/MS-analysis). Reference incubations were performed without NADPH. Liquid-liquid-extraction of the assay with diethylether was accomplished after addition of internal standard, reaction termination by acetonitrile and acidification. The separated organic layer was concentrated under a stream of nitrogen and the residue was dissolved in mobile phase.

## LC-Analysis:

As the activated ketone moiety of **1** exhibits hydrate formation in aqueous solutions, besides the peak of **1** a peak for its hydrate form could be detected during RP-chromatography (see figure 6). Hence, a normal-phase chromatography with isohexane/THF was employed.

A 50  $\mu$ l volume of the resolved assay residue was injected onto Lichrospher 100-5 CN (250 x 4 mm, pre column 4x4 mm) column (Merck, Darmstadt) at a flow rate of 0.75 ml/min, column oven temperature of 25 °C and a UV detection wavelength of 240 nm. The mobile phase consisted of isohexane / THF 92:8 (0.1% TFA) (A) and isohexane / THF 50:50 (0.1% TFA) (B). The following gradient was applied (A%): 0 min: 90, 15 min: 20, 18 min: 20, 20 min: 90, 27 min: 90.

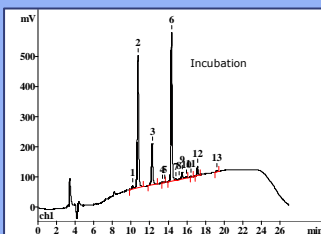


Figure 1

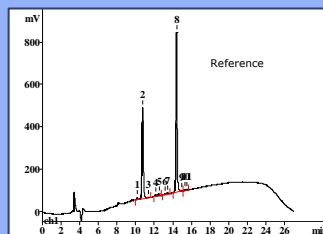


Figure 2

## Results:

The chromatogram of the reference (figure 2) displays the internal standard (peak no. 2: 10.8 min) and the analyt peak of **1** (peak no. 8: 14.4 min) as main peaks. The incubation (figure 1) resulted in the formation of metabolites (peak no. 3: 12.3 min, no. 9: 15.5 min, no. 12: 17.2 min). Peak 3 was the main metabolite with about 18% of the peak area ratio of **1** in the reference assay. Approximately 69% of **1** were not metabolised within 30 min.

After spiking the incubation probe with substance **2**, it could be assumed that this compound was the main metabolite.

## LC/MS<sup>n</sup>-Analysis:

LC/MS<sup>n</sup> was employed to verify the alcohol **2** as the main metabolite.

Since LC/MS investigations with normal-phase chromatography were not successful, a RP-chromatographic method had to be developed.

A 100  $\mu$ l volume of the dissolved assay residue was injected onto Kromasil 100-5 C18 (60 x 2 mm, inletfilter 2  $\mu$ m) column (CS, Langerwehe) at a flow rate of 0.4 ml/min and a UV detection wavelength of 235 nm. The mobile phase consisted of 20mM ammonium formiate / acetonitrile 80:20 (A) and 20mM ammonium formiate / acetonitrile 15:85 (B). The following gradient was applied (A%): 0 min: 55, 10 min: 0, 11 min: 0, 12 min: 55, 15 min: 55

Two minutes after injection, the MS-detector (LCQ<sup>®</sup> Thermo-Finnigan) was activated and structure determination was performed using ESI-MS<sup>n</sup> in negative mode.

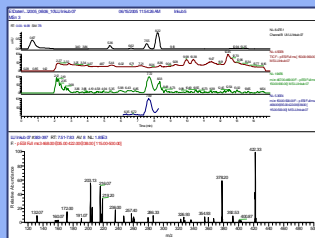


Figure 3

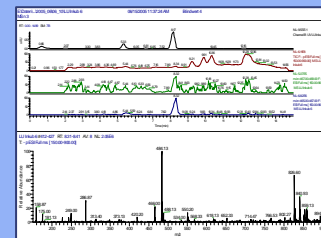


Figure 4

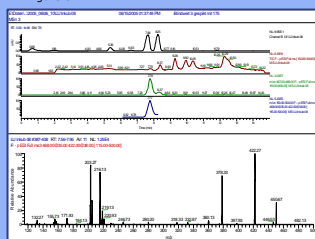


Figure 5

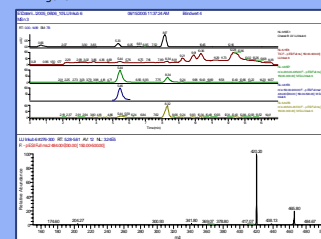


Figure 6

## LC/MS<sup>n</sup> results:

The alcohol **2** was confirmed as the main metabolite with rat liver microsomes. Representative ESI-MS and MS<sup>3</sup> data of the incubation assay are shown in figure 3, which reveal a formiate adduct of **2** at  $m/z$  468 [M+45]<sup>-</sup>. After isolation and fragmentation of  $m/z$  468 the daughter ion  $m/z$  422 [M-H]<sup>-</sup> fragmentation pattern exhibits MS/MS product ions at  $m/z$  378, 216 and 203. The MS<sup>3</sup>-spectrum of a reference probe spiked with **2** (figure 5) corresponds to latter fragmentation pattern. The MS<sup>2</sup> data of the hydrate of **1** is shown in figure 6. The formiate adduct [M+18+45]<sup>-</sup> at  $m/z$  484 was isolated and the following fragmentation revealed **1** as the product ion at  $m/z$  420 [M-H]<sup>-</sup>. Figure 4 represents a full scan of **1** in the range of 8.31-8.41 min.

## Literature:

- [1] Ludwig J., *PhD Thesis, University of Münster (2004)*
- [2] Ansele, J. H.; Thakker, D. R., *J Pharm Sci* **2004**, 93, (2), 239-55.
- [3] Zuber, R.; Anzenbacherova, E.; Anzenbacher, P., *J Cell Mol Med* **2002**, 6, (2), 189-98
- [4] Cottrell, L.; Golding, B. T.; Munter, T.; Watson, W. P., *Chem Res Toxicol* **2001**, 14, (11), 1552-62.
- [5] Saito, K.; Kim, H. S.; Sakai, N.; Ishizuka, M.; Kazusaka, A.; Fujita, S., *J Pharm Sci* **2004**, 93, (5), 1271-8.

## Acknowledgement:

Financial support by Merckle GmbH is greatly appreciated.