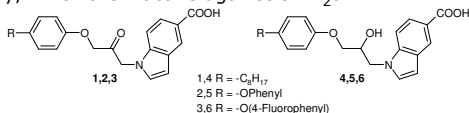


## Introduction:

Inhibitors of cytosolic phospholipase A<sub>2</sub>α (cPLA<sub>2</sub>α) are assumed to provide a novel therapeutic with application in many inflammatory diseases [1]. Certain 1-indol-1-ylpopan-2-ones, such as compounds **1** [2] and **2** [3], are potent inhibitors of cPLA<sub>2</sub>α.

An important part of the pharmacophore of these substances is the activated electrophilic ketone moiety, which is supposed to form covalent binding interactions with a serine of the active site of cPLA<sub>2</sub>α. In earlier investigations metabolic reduction of the ketone by rat liver microsomes led to the alcohols **4** [4] and **5**, respectively, which are inactive against cPLA<sub>2</sub>α.



As inter-species differences and variations of CYP isoforms are known [5] we now investigated, which human CYP450 isoenzymes are responsible for metabolism of compound **2** by inhibitor approach [6].

## Incubation procedure:

2 µl of analyte solution (final concentration 25 µM) and -where required- 1 µl inhibitor solution dissolved in DMSO were added to potassium phosphate buffer (100 mM, pH 7.4 / 3 mM MgCl<sub>2</sub>) containing 20 pmol BD Supermix™ cDNA-expressed CYP450 isoenzymes (1A2, 2C8, 2C9, 2C19, 2D6 and 3A4) cocktail to give a final volume of 200 µl. The reaction was started by the addition of 50 µl NADPH solution (15 mM) and carried out at 37 °C in a shaking water bath for 2h. Reference incubations were performed without NADPH. The incubations were stopped by the addition of ice-cold 375 µl MeCN with 0.5 % HCOOH (v/v). The samples were centrifuged and the supernatants were analysed. The analyte solution were shown to be stable at 10 °C for 24 h.

For LC-MS investigations 250 µl of ice-cold MeCN was used as stop solution. After acidification with 625 µl of 0.1 M phosphoric acid liquid-liquid-extraction of the samples with 2 x 2.5 ml diethylether was accomplished. The separated organic layers were concentrated under a stream of nitrogen and the residue was reconstituted in 300 µl mobile phase.

Inhibitor	CYP450 Isoenzymes	Inhibition Literature (%)	Assay concentration (µM)
β-Phenylthiopyline	1A2	6,7	20
Quercetin hydrate	2C8	6,2	10/50/100/300
Sulfaphenazole	2C9	0,4	20
Tranylcypromine	2C19	12,5	125
Quinidine	2D6	0,030	2
Ketoconazole	3A4	0,04	2
Lipoic acid	NADPH-Reductase	5000	

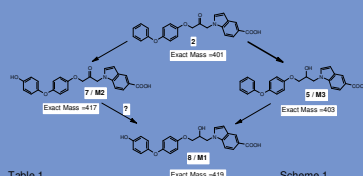


Table 1

Scheme 1

## LC-Analysis:

A 100 µl of sample was injected onto Kromasil 100-5 ODS (150 x 3.2 mm, with securityguard™cartridge) column (Phenomenex) at a flow rate of 0.6 ml/min, column oven temperature of 30 °C and a UV detection wavelength of 235 nm. The mobile phase consisted of MeCN / 20 mM ammonium formate 20:80 (0.2% HCOOH) (A) and MeCN / 20 mM ammonium formate 85:15 (0.2% HCOOH) (B). The following gradient was applied (A%): 0 min: 90, 23 min: 10, 24 min: 10, 25 min: 90, 35 min: 90.

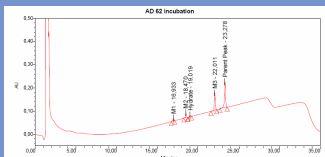


Fig. 1

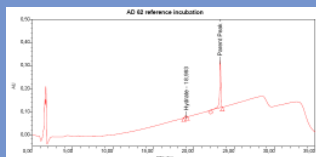


Fig. 2

## Results - Metabolism:

The chromatogram of the reference incubation (Fig. 2) displays the hydrate [4] and the analyte peak of **2**. The incubation (Fig. 1) resulted in the formation of metabolites (**M1**: 16.9 min, **M2**: 18,5 min, **M3**: 22,0 min, Scheme 1). M 3 was the main metabolite with about 30 % relative peak area. M2 was formed by about 10 % and M1 derived out of **5** at circa 6 %. Approximately 53 % of **2** were not metabolised within 120 min.

After spiking the incubation probe with substance **5**, it could be assumed that this compound was the main metabolite.

## Results - Inhibition Investigation:

Inhibition assays (Table 1) revealed CYP 2C19 and 2C8 as the main metabolic pathways of **2** (Scheme 1). Quercetin inhibited the reduction of **2** and in minor the hydroxylation of the phenyl-function. Tranylcypromine exclusively avoided the hydroxylation of the aryl-group of **2** and **5**, respectively. As the 4'-fluorophenyl-derivate **3** did not show hydroxylation (Fig. 6), the 4'-position can be assumed as the preferred reaction site. CYP 3A4 is involved in hydroxylation reaction inferior.

## LC/MS<sup>n</sup>-Analysis:

LC/MS<sup>n</sup> and counter synthesis were employed to verify the metabolites of **2** and **3**, respectively.

A 60 µl (70 µl at LC/MS<sup>3</sup>) volume of reconstituted assay residue was injected onto Kromasil 100-5 ODS (150 x 3.2 mm, with securityguard™cartridge) column (Phenomenex) at a flow rate of 0.6 ml/min and a UV detection wavelength of 235 nm. The mobile phase consisted of MeCN / 20 mM ammonium formate 20:80 (A) and MeCN / 20 mM ammonium formate 85:15 (B). The following gradient was applied (A%): 0 min: 90, 23 min: 0, 24 min: 0, 25 min: 90, 35 min: 90. Three minutes after injection, the MS-detector (LCQ® Thermo-Finnigan) was activated and structure determination was performed using ESI-MS<sup>n</sup> in negative mode.

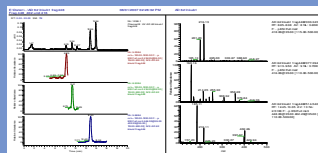


Fig. 3

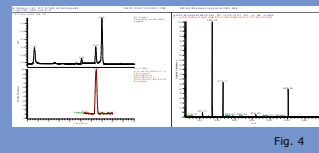
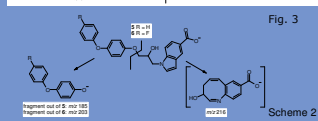


Fig. 4



Scheme 2

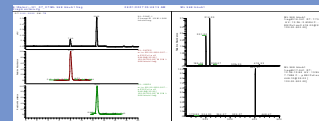


Fig. 5

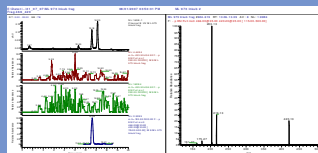


Fig. 6

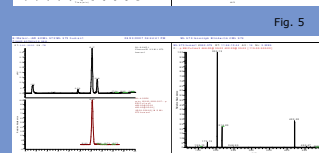


Fig. 7

## Results - LC/MS<sup>n</sup>

LC/MS<sup>n</sup> investigations of the metabolism of **2** confirm the alcohol **5** (**M3**) as the main and **7/8** (**M2/M1**) as minor metabolites (Scheme 1). Representative ESI-MS<sup>2/3</sup> data of the incubation assay are shown in Fig. 3. Isolation and fragmentation of the formate adduct of **5** at *m/z* 448 [M+45]<sup>-</sup> yields the daughter ion at *m/z* 402 [M-H]<sup>-</sup>, which fragments mainly to MS/MS product ions at *m/z* 216 and 185. The assumed structures are represented in Scheme 2. The MS<sup>3</sup>-spectrum of a reference probe spiked with **5** (Fig. 4) corresponds to latter fragmentation pattern. The MS<sup>2</sup> data of **8** (*m/z* 418) (Fig. 3) shows fragments at *m/z* 216 and 201. The product ion *m/z* 201 illustrates the hydroxylation at the phenyloxyphenyl-part. **7** can be detected at *m/z* 416.

Incubation of **5** forms the expected metabolite **8**, which was verified by the fragmentation pattern (Fig. 5).

Incubation of the 4'-fluorophenyl-derivate **3** yields **6** (*m/z* 420) and no further hydroxylation can be observed (Fig. 6). The fragmentation pattern of a spiked reference probe confirms **6** as the main metabolite (Fig. 7).

## Literature:

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- [3] Drews A. et al., unpublished results
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- [6] Bourrie M. et al., *J Pharmacol Exp Ther.* **1996**, 277, 321-32